

# **RESEARCH ARTICLE**

## *In-vitro* Assessment of Fungicides and pH Levels on the Mycelial Growth and Sclerotia Production of *Sclerotium rolfsii* Sacc. causing White Rot of Onion in Manipur

Rimamay Konjengbam<sup>1\*</sup>, Naorem Iboton Singh<sup>1</sup> and Rajkumari Tombisana Devi<sup>2</sup>

<sup>1</sup>Department of Plant Pathology, College of Agriculture, Central Agricultural University, Imphal West, Manipur, India <sup>2</sup>Department of Plant Pathology, College of Post Graduate Studies in Agricultural Sciences, Central Agricultural University, Barapani, Meghalaya, India

## ABSTRACT

An onion white rot fungicide, Sclerotium rolfsii Saccardo, was tested in Manipur. The fungicides were tested on PDA using the poisoned food approach. The effect of pH on potato dextrose broth was assessed by weighing dry mycelium and counting sclerotia generated by the fungus. The white rot pathogen was completely inhibited by three fungicides: Hexaconazole (500 and 1000ppm), Propiconazole (500 and 1000ppm), and Mancozeb (1000 and 2000ppm). S. rolfsii produced 0.62g dry mycelial weight and 1069 sclerotia in pH 4.0. At pH 5.0, the greatest mycelial growth was 0.76g, followed by 0.72g at pH 6.0. The most sclerotia were generated at pH 7.0, followed by PH 6.6 and 6.0.

### Article history:

Received: January 29, 2021 Accepted: March 11, 2021 Published: March 25, 2021 Keywords: Sclerotium rolfsii; White rot; Onion; Fungicides; pH

\*Corresponding author e-mail address: rimamay24@gmail.com (Konjengbam. R)

#### **INTRODUCTION**

Onion is an cool-season crop and belongs to family *Amaryllidaceae*. Onion is known for its high nutritional value containing water (89.11g), protein

(1.1g), carbohydrate (0.34g), fiber (1.7g), total sugar (4.24g), vitamin C (74mg), potassium(146mg), phosphorus (29mg), calcium (23mg), magnesium (10mg) and sodium (4mg) per 100g of onion bulb (Pareek et al., 2017). It has a rich source of various

organo-sulphur compounds. These compounds are responsible for their flavour (Mahajan et al., 2017). Onion contains a range of oxidative enzymes, namely, quercetin, S alkaneylcystiene sulfoxide, superoxide dismutase, catalase, peroxidise and glutathione peroxidase (Stajner and Varga, 2003; Dini et al., 2008; Nile et al., 2017).

In Manipur, onion suffers from various diseases. White rot of onion caused by Sclerotium rolfsii Saccardo is a severe disease that has devastated onion cultivation. The disease has been observed at commercial onion growing locations in the valley districts of Manipur. The disease becomes evident only late in the crop season, near harvesting period, when the leaves start yellowing, drooping, blighting and wilting (Mathur and Sharma, 2002). The harvested onion bulbs are soft, watery and rots accompanied by white cottony mycelium and small white, brown and black colour sclerotia. The fungus has a wide range of hosts, including cereals, solanaceous crops, legumes, crucifers, cucurbits, flowers and even weeds (Aycock, 1966; Punja, 1985; Punja, 1988). The fungus is well known for producing oxalic acid (Punja and Jenkins, 1984; Paramsivan et al., 2013) in addition to several enzymes, including endo-polygalacturonase, endo-pectimethyl polygalacturonase and cellulase (Punja et al., 1985). The fungus produces sclerotia as resting structure for its survival (Punja, 1985; Xu et al., 2008). This pathogen also survives as dormant mycelium in infected plants and attacks plants' collar region (Mullen, 2001). The sclerotial wall contains melanin pigment and a considerable amount of nonhydrolysable residue, lipids, and ash, making sclerotia tolerant to biological and chemical degradation (Chet et al., 1967). There is also variability in morphology, cultural, physiology and pathogenicity among isolates of Sclerotium rolfsii (Sarma et al., 2002; Shukla and Pandey, 2008; Kumar et al., 2014). Therefore, the management of S. rolfsii causing white rot of onion in Manipur with fungicides that inhibit mycelial growth, as well as sclerotia production, is very crucial because the fungus is soilborne, polyphagus and the sclerotia remain viable for a long period (Punja; 1985, Kator et al., 2015).

## **MATERILAS AND METHODS**

Soaked in 1 percent sodium hypochloride solution for 10 minutes, the infected onion bulbs were recovered. PDA was prepared and fungicides were added to 50ml molten PDA, mixed well, and placed onto three sterilised petriplates. A 90mm petriplate was filled with molten PDA media and allowed to cool. The sterilised pieces were dried and inoculated on potato dextrose agar for 4 days at 28oC. The pathogen was purified by hyphal tip cut and re-isolated on PDA solid state. Throughout the research time, the pathogenic culture was kept on PDA and sub-cultured to fresh PDA.

The three non-systemic fungicides (Captan, Mancozeb, and Blitox) and four systemic fungicides (Hexaconazole, Thiophanate methyl, Propiconazole, and Carbendazim) were used against Sclerotium rolfsii (Grover and Moore, 1962). The same sterilised PDA medium was used to test the efficiency of several fungicides. Each replication had three petriplates with a sufficient control San fungicide. Each treatment had three replications. A three-day-old culture of the fungus was injected on each plate after proper PDA solidification. The plates were incubated at 28oC inverted until the test fungus had fully grown on the control plates. Using Vincent's approach, we estimated the percent inhibition of mycelial growth (1927).

After 15 days of incubation, sclerotia generated by two fungicide concentrations were counted. Using Vincent's approach, we estimated the percent suppression of sclerotia generation (1927).

This study examined the influence of pH on growth and formation of sclerotia in the fungus. Each pH level had three replications, including control. Sterilised conical flasks with adjusted potato dextrose broth at 1210C for 20 minutes at 15lbs. Using a three-day-old potato dextrose agar culture, each conical flask was infected and incubated at 2810C. The weight of each filter paper was noted beforehand. The separate potato dextrose broths were filtered through preweighed filter papers and allowed to dry fully. After 15 days, the dried mycelial development of the fungus was estimated.

### **RESULTS AND DISCUSSION**

Morphological and taxonomic data from the monographs helped identify the fungus as Sclerotium rolfsii Saccardo (Saccardo, 1913; Mordue, 1974; Punja, 1985).

Among the studied fungicides, Hexaconazole, Propiconazole, and Mancozeb were effective against Sclerotium rolfsii, inhibiting mycelial growth and sclerotia development. All fungicides' efficacy increased with concentration. At 500ppm and 1000ppm, thiophanate methyl inhibited mycelial growth and sclerotia formation by 82.22 and 100%, respectively. Copper oxycholride and Captan are inferior non-systemic fungicides to Mancozeb. Carbendazim is inferior to other systemic fungicides in reducing mycelial growth and sclerotia formation. Both Copper oxychloride and Captan concentrations could generate sclerotia. Johnson et al. (2008) showed that 1000ppm Hexaconazole and Propiconazole totally inhibited Sclerotium rolfsii growth. In the study, Rakholiya et al. (2015) found that Hexaconazole and Propiconazole at 500ppm and Mancozeb at 1000ppm totally suppressed mycelial development and sclerotia generation of S. rolfsii (groundnut stem rot).

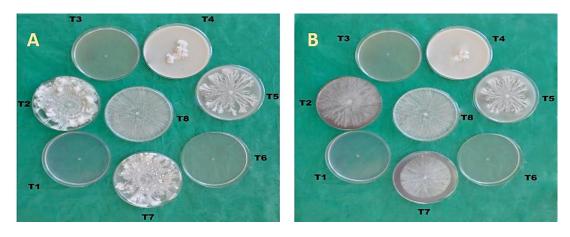
Propionazole 500ppm and 1000ppm and Mancozeb 1000ppm and 2000ppm showed 100% suppression of mycelial growth according to Rangarani et al. 1000ppm carbendazim was less effective. S. rolfsii isolates were entirely inhibited by Hexaconazole at 500ppm, whereas Mancozeb at 1000ppm and 2000ppm completely inhibited select isolates, according to Sharma and Dhruj (2018). Carbendazim and Captan were less inhibitory at 0.1 percent (1000ppm) concentration, according to Bhagat and Chakraborty (2013). According to Mahato et al. (2014 S. rolfsii and had no influence on mycelial growth. Arunasri et al. (2011), Manu et al. (2012), Deepthi (2014), Suneeta et al. (2017), Shirsole et al. (2019) and Sahana et al (2020).

**Table 1**. Effect of fungicides at first concentration

S.No.	Fungicides	Concentration	Growth	Inhibition	Sclerotia	Per cent inhibition
		(ppm)	(cm)*	(%) over	production*	on sclerotia
				control		production
T1	Hexaconazole (5%	500	0.00	100	0	100
	EC)		(0.71)			
T2	Copper oxychloride	1000	8.83	1.88	113	47.19
	(50% WP)		(3.05)			
Т3	Mancozeb	1000	0.00	100	0	100
	(75% WP)		(0.71)			
T4	Thiophanate methyl	500	1.60	82.22	0	100
	(70% WP)		(1.43)			
Т5	Captan	1000	7.75	13.88	106	50.46
	(50% WP)		(2.84)			
Т6	Propiconazole	500	0.0	100	0	100
	(25% EC)		(0.71)			
Τ7	Carbendazim	500	8.33	7.44	109	49.06
	(50% WP)		(2.97)			
Τ8	Control	-	9.00	0.00	214	0.00
			(3.08)			
	SE(d) ±	-	0.34	-	-	-
			(0.08)			
	CD (0.05)	-	0.74	-	-	-
			(0.19)			

Sl. No.	Fungicides	Concentration	Growth	Inhibition	Sclerotia	Per cent inhibition
		(ppm)	(cm)*	(%) over	production*	on sclerotia
				control		production
T1	Hexaconazole (5% EC)	1000	0.00	100	0	100
			(0.71)			
T2	Copper oxychloride	2000	8.67	3.66	47	78.73
	(50% WP)		(3.03)			
Т3	Mancozeb	2000	0.00	100	0	100
	(75% WP)		(0.71)			
T4	Thiophanate methyl	1000	0.93	89.66	0	100
	(70% WP)		(1.20)			
Т5	Captan	2000	6.53	27.44	57	71.94
	(50% WP)		(2.65)			
Т6	Propiconazole	1000	0.0	100	0	100
	(25% EC)		(0.71)			
Τ7	Carbendazim	1000	7.23	19.66	62	71.94
	(50% WP)		(2.78)			
Τ8	Control	-	9.00	0.00	221	0.00
			(3.08)			
	SE(d) ±	-	0.08	-	-	-
			(0.01)			
	CD (0.05)	-	0.19	-	-	-
			(0.04)			

Table 2. Fungicides effects at second concentration



**Figure 1.** Fungicides effect on the mycelial growth of *S. rolfsii*at first (A) and second concentration (B). T1, Hexaconazole; T2, Copper oxychloride; T3, Mancozeb; T4, Thiophanate methyl; T5, Captan; T6, Propiconazole; T7, Carbendazim; T8, Control

Sclerotium rolfsii grows in all pH tested, 4.0 to 7.0. The pH of the control potato broth was found to be 6.6. S. rolfsii grew to 0.76g at pH 5.0. From pH 4.0 to 7.0, sclerotia were developed. Sclerotia (1109) were generated at pH 7.0. In pH 4.0, mycelial growth (0.62g) and sclerotia development (1069) were

minimal. Richhariya (1984) showed that pH 5.0 was optimal for S. rolfsii mycelial development. Many other authors have observed that S. rolfsii can grow in a wide range of pH. The present findings agree with Muthukumar and Venkatesh (2013). They found that pH 5.0 produced the most mycelial dry weight, followed by pH 6.0. According to Chaurasia et al. (2013), pH 4.0 to 7.0 is optimal for sclerotia

formation. Sarker et al. (2013) found a pH range of 4.5 to 6.5 for dry mycelial growth of S. rolfsii. According to Zape et al. (2013), sclerotia numbers peak at pH 7.0. They found that S. rolfsii grew best at pH 4–7 (Ayed et

al. 2018). Kushwaha et al. (2019) reported good sclerotia development at pH 6.0, 6.5, and 7.0. Similarly, Sri et al. (2020) found that pH 7.0 favoured S. rolfsii sclerotia production.

pН	Mycelial dry weight(g) *	Sclerotia production*
4	0.62	1069
4.5	0.70	1070
5.0	0.76	1075
6.0	0.72	1089
7.0	0.60	1109
Control (6.6)	0.64	`1093
$SE(d) \pm$	0.03	-
CD(0.05)	0.07	-

**Table 3.** Effect of different pH levels on the mycelial growth



Figure 2. The effect of various levels of pH on the mycelial growth

## CONCLUSION

The present investigations disclosed that *S. rolfsii* causing white rot of onion in Manipur can grow and produce sclerotia in a broad range of pH levels. Among the fungicides tested, Hexaconazole, Propiconazole and Mancozeb were effective against the pathogen, and completely inhibited both mycelial growth and sclerotia production. The fungicides effective against this fungus under laboratory conditions should further be tested for their efficacy

under field conditions. Since *Sclerotium rolfsii* is soilborne and survives as mycelium or sclerotia, fungicides that inhibit both mycelial growth and sclerotia production can be employed in integrated disease management strategies.

## REFERENCES

- Arunasri, P., Chalam, T. V., Eswara, R. N. P., Tirumala, R., & Ravindra, R. B. (2011). Investigations on fungicidal sensitivity of *Trichoderma* spp. and *Sclerotium rolfsii* (collar rot pathogen) in crossandra. *International Journal* of Applied Biology and Pharmaceutical Technology, 2(2), 290-293.
- Aycock, R. (1966). Stem rot and other diseases caused by *Sclerotium rolfsii*. N.C State University Technical Bulletin, 174, p202.
- Ayed, F., Jabnoun-Khiareddine, H., Aydi-Ben-Abdallah, R., & Daami-Remadi, M. (2018). Effects of pH and Aeration on Sclerotium rolfsii Sacc. Mycelial Growth, Sclerotia Production and Germination. International Journal of Phytopathology, 7(3):111-121. <u>https://10.33687/phytopath.007.03.2688</u>.
- Bhagat, I., & Chakraborty, B. (2013). Efficacy of fungicides against sclerotial blight of tea plant. *Our Nature*, *11*(30), 208-210.
- Chet, I., Henis, Y. & Mitchell, R. (1967). Chemical composition of hyphal and sclerotial walls of *Sclerotium rolfsii* Sacc. *Canadian Journal of Microbiology*, *13*(2),137-141.
- Deepthi, K. C. (2014). In vitro Evaluation of Fungicides Against *Sclerotium rolfsii*Sacc causing Stem Rot of Groundnut. *International Journal of Scientific Research*, 3(12),1-2.
- Chowdary, K. A., Reddy, D. R., & Rao, K. C. (1998). Efficacy of systemic (triazoles) and non-systemic fungicides against sclerotial wilt of bell pepper caused by *Sclerotium rolfsii* Sacc. *Indian Journal of Plant protection*, 26(2),125-130.
- Dini, I., Tenore, G. C., & Dini, A. (2008). S-Alkenyl Cysteine Sulfoxide and Its Antioxidant Properties from Allium cepa var. tropeana (Red Onion) Seeds. Journal of Natural Products, 71(12), 2036-2037. doi: 10.1021/np800237w
- Grover, R. K., & Moore, J.D. (1962). Toxicometric studies of fungicides against brown rot organism, *Sclerotinia fruiticola* and S. laxa. *Phytopathology*, *52*, 876-880.
- Johnson, M., & Subramanyam, K. (2000). *In-vitro* efficacy of fungicides against stem rot pathogen (*Sclerotium rolfsii*) of groundnut. *Annals of Plant Protection Sciences*, 8(2), 255-257.
- Johnson, M., Reddy, P. N., & Reddy, D.R. (2008). Comparative efficacy of rhizosphere mycoflora, fungicides, insecticides and herbicides against groundnut stem rot caused by *Sclerotium rolfsii*. *Annals of Plant Protection Sciences*, *16*(2),414-418.
- Kator, L., Hosea, Z. Y., & Oche, O.D. (2015). *Sclerotium rolfsii;* Causative organism of southern blight, stem rot, white mold and sclerotia rot disease. *Annals of Biological Research*, *6*(11),78-89.
- Kumar, R., Santhoshi, M.V. M., Krishna, T. G., & Reddy, K.R. (2014). Cultural and Morphological Variability Sclerotium rolfsii Isolates Infecting Groundnut and Its Reaction to Some Fungicidal. *International Journal of Current Microbiology and Applied Sciences*, 3(10), 553-561.
- Kushwaha, S. K., Kumar, S., Chaudhary, B., & Sahu, R. (2019). Effect of Different Media, pH and Temperature on Growth and Sclerotia Formation of *Sclerotium rolfsii* Sacc. causing Collar rot of Lentil. *Chemical Science Review and Letters*, 8(29),1-5.
- Mahajan, V., Ghodke, P., Bhagat, K. P., Kalyani, G. Soumia, P. S., Shirsat, D., & Singh, M. (2017). Use of Onion (*Allium cepa* L.) as Medicine. *International Journal of Noni Research*, *12*(1-2), 39-43.
- Mahato, A., Mondal, B., Dhakre, D. S., & Khatua, D. C. (2014). *In vitro* sensitivity of *Sclerotium rolfsii* towards some fungicides and botanicals. *Scholars Academic Journals of Biosciences*, 2(7), 467-471.
- Manu, T. G., Nagaraja, S., Chetan, S., & Hosamani, J. V. (2012). Efficacy of Fungicides and Biocontrol Agents against Sclerotium rolfsii causing Foot Rot Disease of Finger Millet, Under In vitro Conditions. Global Journal of Agriculture, Biology and Health Sciences, 1(2), 46-50.
- Mathur, K., & Sharma, S. N. (2002). Bulb rot of onion induced by *Sclerotium rolfsii* a new threat to onion cultivation in Rajasthan. *Journal of Mycology and Pant Pathology*, *32*(1), 132-133.
- Mordue, J. E. M. (1974). *Sclerotium rolfsii.* CMI descriptions of pathogenic fungi and bacteria. No. 410. Commonwealth Mycological Institute, Kew, Surrey, England.

- Mullen, J. (2001). Southern blight, Southern stem blight, White mold. *The Plant Health Instructor*. https://doi.org/10.1094/PHI-I-2001-0104-01
- Muthukumar, A., & Venkatesh, A. (2013). Physiological studies of *Sclerotium rolfsii* Sacc. causing collar rot of peppermint. *African Journal of Plant Pathology*, *12*(49), 6837-6842.
- Narasimhan, R. (1969). Physiological studies on the genus sclerotium. I Effect of initial H. Ion concentration on the growth of *Sclerotium rolfsii* and *sclerotium oryzae* in different inorganic nitrogen media. *Indian Phytopathology*, *22*, 115-123.
- Nile, S. H., Nile, A. S., Keum, Y. S. & Sharma, K. (2017). Utilization of quercetin and quercetin glycosides from onion (*Allium cepa* L.) solid waste as an antioxidant, urease and xanthine oxidase inhibitors. *Food Chemistry*, 235(15), 119-126, <u>https://doi.org/10.1016/j.foodchem.2017.05.043</u>
- Paramsivan, M., Mohan, S., Muthukhrishnan, N. & Chandrasekaran, A. (2013). Degradation of oxalic acid (OA) producing *Sclerotium rolfsii* (Sacc.) by organic biocides. *Archives of Phytopathology and Plant Protection*, 46(3), 357-363. <u>https://doi.org/10.1080/03235408.2012.740983</u>
- Pareek, S., Sagar, N. A., Sharma, S., & Kumar, V. (2017). Onion (*Allium cepa* L.): Chemistry and Human Health. In E.M. Yahia, *Fruit and Vegetable Phytochemicals: Chemistry, Nutrional Value and Stability* (2<sup>nd</sup> ed., pp.1145-1162). John Wiley and Sons, Ltd.
- Punja, Z. K. (1985). The biology, ecology, and control of *Sclerotium rolfsii*. *Annual Review of Phytopathology*, *23*, 97-127.
- Punja, Z. K., & Jenkins, S. F. (1984). Influence of medium composition on mycelial growth and oxalic acid production in *Sclerotium rolfsii*. *Mycologia*, *76*, 947-950.
- Punja, Z. K., Huang, Z. S., & Jenkins, S.F. (1985). Relationship of Mycelial Growth and Production of Oxalic Acid and Cell Wall Degrading Enzymes to Virulence in *Sclerotium rolfsii. Canadian Journal of Plant Pathology*, 7(2), 109-117.
- Punja, Z. K. (1988). *Sclerotium (Athelia) rolfsii*, a pathogen of many plant species. In G.S. Sindhu, Advances in plant pathology (pp. 523-534). Academic Press.
- Rangarani, A., Rajan, C.P.D., Harathi, P.N., Bhaskar, B., & Sandhya, Y. (2017). Evaluation of Fungicides and Herbicides on *Sclerotium rolfsii*, Incitant of Stem Rot Diseases in Groundnut. *International Journal of Pure & Applied Bioscience*, *5*(3), 92-97.
- Rakholiya, K. B. (2015). Screening of fungicides against *Sclerotium rolfsii* causing stem rot of groundnut. *An International Quarterly Journal of Life Sciences, 10*(2), 691-694.
- Saccardo, P.A. (1913). Sclerotium rolfsii. Sylloge Fungorum XXII, Pavia, Italy.
- Sahana, B., Manjunatha Reddy, T. B., Mushrif, S. K., Anjaneya Reddy, B. & Doddabasappa, B. (2020). Evaluation of Fungicides against Stem Rot of Capsicum caused by *Sclerotium rolfsii* Sacc. *International Journal of Chemical Studies*, 8(4), 306-312.
- Sarker, B. C., Adhikary, S. K., Sultana, S., Biswas, A., & Azad, S. F. D. (2013). Influence of pH on growth and sclerotia formation of *Sclerotium rolfsii* causal agent of foot rot disease of betelvine. *IOSR Journal of Agriculture and Veterinary Science*, 4(1), 67-70.
- Sarma, B. K., Singh, U. P., & Singh, K. P. (2002). Variability in Indian isolates of *Sclerotium rolfsii. Mycologia*, 94(6),1051-1058. <u>https://doi.org/10.2307/3761870</u>.
- Sharma, S.L. & Kaushal, B.R. (1979). Cultural and physiological studies with sunflower isolate of *Sclerotium rolfsii*. *Indian journal of the Mycology and Plant Pathology*, *9*, 105-107.
- Sharma, R. K., & Dhruj, I. U. (2018). In vitro Evaluation of Some Fungicides against Indian Isolates of Sclerotium rolfsiiSacc. [Teleomorph: Atheliarolfsii (Curzi) Tu & Kimbrough]. International Journal of Current Microbiology and Applied Sciences, 7(11), 1561-1586.
- Shirsole, S., Khare, N., Lakpale, N., & Kotasthane, A. (2019). Evaluation of fungicides against *Sclerotium rolfsii*Sacc. Incitant of collar rot of chickpea. *The Pharma Innovation Journal*, *8*(12), 310-316.
- Shukla, R., & Pandey, A. K. (2008). Pathogenic diversity of *Sclerotium rolfsii* isolates, a potential biocontrol agent against *Parthenium hysterophorus* L. *African Journal of Environmental Science and Technology*, 2(5)-124-126.

- Sri, P. T., Sajeena, A., Johnson, J. M., John, J., & Radhika, N. S. (2020). Factors influencing the incidence of basal stem rot and blight disease caused by *Sclerotium rolfsii* in vegetable cowpea and its management using botanicals. *Journal of Biological Control*, 34(3), 215-222.
- Suneeta, P., Aiyanathan, E.A. & Sevugapperumal, N. (2017). Evaluation of *Trichoderma* spp. and Fungicides in the Management of Collar Rot of Gerbera Incited by *Sclerotium rolfsii*. *Journal of Pure and Applied Microbiology*, 11(2),1161-1168.
- Vincent, J. M. (1927). Distortion of fungal hyphae in presence of certain inhibitors. Nature, 159:850

Walker, J. C. (1924). White rot of Allium in Europe and America. *Phytopathology*, 14(7), 315-322.

- Xu, Z., Gleason, M. L., Mueller, D. S. & Esker, P. D. (2008). Overwintering of Sclerotium rolfsii and S. rolfsii var. delphinii in Different Latitudes of the United States. Plant Disease, 92(5), 714-724. doi: 10.1094/pdis-92-5-0719.
- Zape, A. S., Gade, R. M. & Ravindra, S. (2013). Physiological studies on different media, pH and temperature on *Sclerotium rolfsii* isolates of soybean. *Scholarly Journal of Agricultural Science*, *2*(6), 238-241.